

INSTRUCTION MANUAL

Mix & G E. coli Transformation Kit & Buffer Set

Catalog Nos. **T3001** & **T3002**

Highlights

- Simple method for generating Mix & G chemically competent E. coli for rapid, reliable and highly efficient DNA transformation (>10⁸ transformants/µg plasmid DNA).
- Straightforward procedure: grow, wash, and then resuspend cells.
- The Mix & G E. coli Transformation Kit (T3001) features a specially formulated **ZymoBroth[™]** growth medium that dramatically increases *E. coli* transformation efficiency, typically on the order of 5 to 100-fold for most lab strains.
- Mix & Go cells can be transformed in seconds without heat shocking, lengthy incubations, or outgrowth steps.

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Product Contents

ZymoBroth™ is included in the *Mix & Go E. coli*Transformation Kit (T3001).

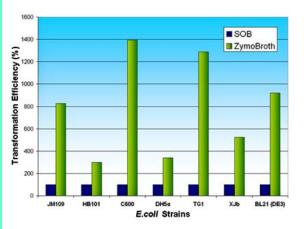
ZymoBroth™ is <u>not</u> included in the *Mis* & G E. coli Transformation Buffer Set (T3002).

	Mix & G E. coli Transformation Kit Includes all buffers for making up to 20 ml Mix & G E. coli. ZymoBroth™ growth medium is included.	Mix & G E. coli Transformation Buffer Set Includes all buffers for making up to 60 ml Mix & G E. coli. ZymoBroth™ growth medium is not included.	Storage Temp.
	T3001 (20 ml)	T3002 (60 ml)	
ZymoBroth™	200 ml	(Not Included)	Room Temp.
Wash Buffer (2X)	10 ml	30 ml	0-8°C
Competent Buffer (2X)	10 ml	30 ml	0-8°C
Dilution Buffer	20 ml	60 ml	0-8°C
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Note - Integrity of kit components is guaranteed for up to one year from date of purchase. Reagents are routinely tested on a lot-to-lot basis to ensure they provide maximal performance and reliability.

Product Description

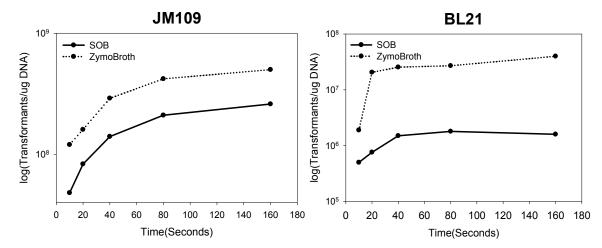
The Mix & G E. coli Transformation Kit & Buffer Set are simple methods for generating Mix & G chemically competent E. coli for rapid, reliable and highly efficient DNA transformation. The methods eliminate the requirement of heat-shocking and related procedures. Instead, transformation can be performed by adding DNA to prepared Mix & G cells and the mixture spread directly to a culture plate. Transformation efficiencies typically range from 10^8-10^9 transformants/ μ g of pUC19 DNA, but can vary depending on the strain of E. coli. Most E. coli strains respond well to the Mix & G preparation method and demonstrate fast transformation kinetics (see figures below).



ZymoBroth™ medium dramatically increases the transformation efficiencies of a broad range of *E. coli* strains. The figure to the left shows the relative transformation efficiencies for different *E. coli* strains generated using SOB and ZymoBroth™. Generally, ZymoBroth™ medium enhances *E. coli* transformation efficiencies better for difficult to transform strains.

Satisfaction of all Zymo Research products is guaranteed. If you should be dissatisfied with this product please call 1-888-882-9682

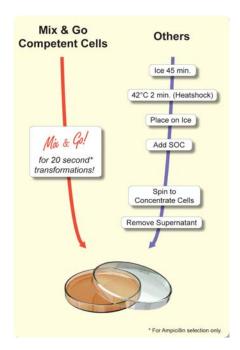
Note -TM Trademarks of Zymo Research Corporation. This product is for research use only and should only be used by trained professionals. Not intended for use in diagnostic procedures. Wear protective gloves and eye protection. Follow the safety diagnostic by your research institution or facility.



Mix & Go E. coli cells prepared with ZymoBroth™ display fast transformation kinetics and high transformation efficiencies. Figures above show the transformation kinetics for JM109 and BL21 strains of E. coli generated using ZymoBroth™ and SOB growth media. PUC19 DNA was used for transformation and the data are the averages of three individual experiments.

The procedures are easy. Simply culture the *E. coli* strain of your choice in **ZymoBroth**™ medium (or SOB), wash and then resuspend the cells in the provided uniquely formulated buffers. The cells are now ready for transformation!

The Min & G E. coli Transformation Kit (T3001) includes all buffers for making up to 20 ml Min & G E. coli from your favorite lab strains. ZymoBroth™ growth medium is included. The Min & G E. coli Transformation Buffer Set (T3002) includes all buffers for making up to 60 ml Min & G E. coli from your favorite lab strains, but ZymoBroth™ growth medium is not included.



Protocol

For Technical Assistance, please contact Zymo Research's Technical Department at 1-888-882-9682 or E-mail tech@zymoresearch.com.

The following procedure is for a 50 ml *E. coli* culture in **ZymoBroth™** (supplied with T3001 only) or SOB medium (see appendix for recipe); however, the volume can be adjusted according to your specific requirements.

Preparation of Mix & Go Cells

1. Use 0.5 ml of fresh, overnight *E. coli* culture grown in LB to inoculate 50 ml **ZymoBroth**™ or SOB medium in a 500 ml culture flask. Shake culture vigorously (150 - 250 rpm) at the appropriate temperature* until the OD_{600nm} is 0.4 - 0.6.

Buffer Preparation Prior to Harvesting the Cells...

The **Wash** and **Competent Buffers** are provided as 2X stock solutions. They need to be diluted to 1X by adding an equal amount of **Dilution Buffer**.

To prepare 5 ml of 1X **Wash Buffer**: Add 2.5 ml **Dilution Buffer** and 2.5 ml of 2X Stock **Wash Buffer**.

To prepare 5 ml of 1X Competent Buffer: Add 2.5 ml Dilution Buffer and 2.5 ml of 2X Stock Competent Buffer.

Please keep these freshly prepared 1X Buffers ice cold. These 1X Buffers are good for 2 days at 0-25°C. It is important that each step of the following procedure should be done on ice or at 0-4°C.

It is important that Steps 2-5 be performed at 0-4°C.

- 2. Transfer the culture from Step 1 to ice. After 10 minutes, pellet the cells by centrifugation at 3,000 3,700 rpm (i.e., 1,600 2,500 x g) for 10 minutes at 0 4°C.
- 3. Remove the supernatant and resuspend the cells gently in 5 ml ice-cold 1X **Wash Buffer**. Re-pellet the cells as in Step 2.
- 4. Completely remove the supernatant and gently resuspend the cells in 5 ml ice-cold 1X **Competent Buffer**.
- 5. Aliquot (on ice) 0.1-0.2 ml of the cell suspension into sterile microcentrifuge tubes. Cells are now ready for transformation with DNA or can be stored below -70°C for transformation at a later time.

Note: The prepared competent cells are referred to as "Min & G" in the procedures that follow.

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^{*} A study by Inou (Gene, 96:23-28, 1990) has shown *E. coli* cells to be highly competent when grown at 18-26°C prior to their preparation. In most cases, this can be achieved by shaking at room temperature for about 10-36 hours. This procedure is covered by US Patent No. 4,981,797 issued to Life Technologies, Inc. No license to use this technology is conveyed expressly or by implication to the purchaser by purchase of the Mix & Gr E. coli Transformation Kit or Mix & Gr E. coli Transformation Buffer Set.

Fast Transformation of Mix & Go Competent Cells*

- Add 1-5 μl plasmid DNA to a tube of thawed Mix & Go cells on ice, mix gently for a few seconds (try to keep the added volume of DNA less than 5% of the total).
- 2. Spread 50-100 µl of the mixture onto a pre-warmed (37°C) culture plate containing Ampicillin. Incubate the plate at the appropriate temperature (e.g., 37°C) for the colonies to grow.

*For Ampicillin selection only. For selection with other antibiotics, see notes Section 4 below.

Notes for High Efficiency Transformation

1. E. coli Strains

Different *E. coli* strains vary in their ability to be transformed with DNA. Strains like JM109, C600, TG1, DH5 α *, XL10 Gold*, and BL21 and its derivatives typically yield the best results when prepared with the Mix & G *E. coli* Transformation Kit.

2. Incubation Time

The "Mix & G" procedure (above) can be used for most transformations using Ampicillin selection and not requiring outgrowth (see Section 4 below). The highest transformation efficiencies can be obtained by incubating Mix & G cells with DNA on ice for 2-5 minutes prior to plating.

3. Prewarming Culture Plates

Chilled plates will decrease *Mix* & *G* cell transformation efficiency. It is recommended that culture plates be pre-warmed to >20°C (preferably 37°C) prior to plating.

4. Addition of SOC Medium to Transformation Mixtures (Outgrowth)

When selecting with Kanamycin, Tetracycline, etc., an outgrowth performed in SOC medium is required for efficient transformation. In most cases, this step can be omitted when selecting with Ampicillin. After the transformation mixture has incubated on ice for 5-10 min, add 4 volumes of SOC (400 μ l of SOC to 100 μ l of transformation mixture) and incubate for 1 hour at 37°C with gentle shaking at 200-300 rpm. Afterwards, spread the mixture directly onto pre-warmed culture plates. Reducing agents [e.g., DTT (Dithiothreitol) and 2-ME (β -mercaptoethanol)] are not required for this procedure.

5. Culture Conditions

E. coli cells become highly competent when cultured at $20-33^{\circ}C$ prior to preparation. Higher temperatures (i.e., $33^{\circ}C-37^{\circ}C$) can decrease the transformation efficiency 2 to 10-fold. Also, cells can be harvested at lower densities (e.g., OD_{600nm} 0.2-0.4) and resuspended in smaller volumes (e.g., 1-3 ml vs. 5 ml as recommended in the standard procedure). Cells harvested at lower densities (OD_{600nm} 0.2-0.6) are usually "more competent" than those cells harvested at higher densities (OD_{600nm} >0.6).

*DH5 α is a trademark of Life Technologies and XL10 Gold is a trademark of Stratagene.

Appendix

Although SOB has traditionally been used for Mix & G cell preparation, **ZymoBroth**TM is now the medium of choice for the generation of Mix & G E. coli that exhibit fast and highly efficient transformation kinetics. However, SOB can still be used in both Mix & G E. coli Transformation Kit and Buffer Set procedures.

SOB Recipe: (1 Liter)

Mix the following ingredients:

20 g Bacto-tryptone 0.58 g NaCl (or 2 ml of 5M NaCl) 10 ml 1M MgCl₂ 5 g Yeast extract 0.19 g KCl (or 0.5 ml 1M KCl) 10 ml 1M MgSO₄

Add ddH₂O to a total volume of 1 liter.

Adjust pH to 6.0-7.0 with NaOH.

Autoclave at 10 psi for 15-20 minutes.

SOC Recipe: (100 ml)

Add 1 ml of a 2 M filter-sterilized glucose solution or 2 ml of 20% (w/v) glucose solution to 100 ml of SOB medium.

References

- Sheridan, P. et al. Phylogenetic Analysis of Anaerobic Psychrophilic Enrichment Cultures Obtained from a Greenland Glacier Ice Core, Appl. Envir. Microbiol., Apr 2003; 69: 2153 – 2160.
- Yokobayashi, Y. et al. From the Cover: Directed evolution of a genetic circuit, PNAS, Dec 2002; 99: 16587 16591.
 Trent, J. et al. A Ubiquitously Expressed Human Hexacoordinate Hemoglobin, J. Biol. Chem., May 2002; 277: 19538 19545.
- Mourez, M. et al. Mapping dominant-negative mutations of anthrax protective antigen by scanning mutagenesis, PNAS, Nov 2003; 100: 13803 –

Ordering Information

Product Description	Catalog No.	Kit Size
Mix & G E. coli Transformation Kit Includes all buffers for making up to 20 ml Mix & G E. coli from your favorite lab strains. ZymoBroth™ growth medium is included.	T3001	Prepare up to 20 ml Competent Cells
Min & G E. coli Transformation Buffer Set Includes all buffers for making up to 60 ml Min & G E. coli from your favorite lab strains. ZymoBroth™ growth medium is not included.	T3002	Prepare up to 60 ml Competent Cells

For Individual Sale	Catalog No.	Amount
ZymoBroth™	M3015-100 M3015-500	100 ml 500 ml
Wash Buffer (2X Stock)	T3001-2-10 T3001-2-30	10 ml 30 ml
Competent Buffer (2X Stock)	T3001-3-10 T3001-3-30	10 ml 30 ml
Dilution Buffer	T3001-4-20 T3001-4-60	20 ml 60 ml

Premade Mix & Go Competent Cells from Zymo Research

Product	Description	Kit Size	Cat No.
JM109 Mix & G Competent Cells	General cloning, blue-white selection, plasmid isolation, healthy strain, transformation efficiency > 10 ⁸ . Genotype: F`traD36 lacl ^q Δ(lacZ)M15 pro A+ B+ / e14 ⁻ (McrA -) Δ(lacproAB)thi gyrA96 (Nal ^r)endA1 hsdR17(r _k ⁻ m _k ⁺)relA1 supE44 recA1	10 x 100 µl aliquots (10 tubes) 96 x 50 µl aliquots (12 x 8-tube strips)	T3003
Zymo 5α Mix & G Competent Cells (Same as DH5α)	General cloning, blue-white selection, plasmid isolation (slow growth, certain plasmids not stable), transformation efficiency > 10^8 . Genotype: $F^-\phi 80 dlac Z \Delta M15 \Delta (lac ZYA-argF)U169 deoR$, recA1 endA1 hsdR17($r_k^-m_k^+$ phoA supE44 λ^- thi-1 gyrA96 relA1	10 x 100 µl aliquots (10 tubes) 96 x 50 µl aliquots (12 x 8-tube strips) 96 x 50 µl aliquots (PCR plates)	T3007 T3009 T3010
HB101 Mix & G Competent Cells	General cloning, plasmid isolation, transformation efficiency > 10 ⁸ . Genotype: F-Δ(gpt-proA)62 leuB6 supE44 ara-14 galkK2 lacY1 Δ(mcrC-mrr) rpsL20 (Str ^r) xyl-5 mtl-1 recA13	10 x 100 µl aliquots (10 tubes)	T3011
C600 Mix & G Competent Cells	General cloning, plasmid isolation, transformation efficiency > 10 ⁸ . Genotype: F ⁻ [e14-(McrA ⁻) or e14 ⁺ (mcrA ⁺] thr- 1leuB6 thi- 1 lacY1 supE44 rfbD1 fhuA21; the original C600 is EcoKr ⁺ m ⁺ McrBC ⁺ (2,3)	10 x 100 µl aliquots (10 tubes)	T3015
TG1 Mix & Go Competent Cells	General cloning, blue-white selection, plasmid isolation, transformation efficiency > 10^8 . Genotype: F'traD36 lacl ^q Δ (lacZ) M15 proA ⁺ B ⁺ /supE Δ (hsdM-mcrB)5 (r_k - m_k - McrB ⁻) thi Δ (lac-proAB)	10 x 100 µl aliquots (10 tubes)	T3017
Zymo 10B Mix & G Competent Cells (Same as DH10B)	For general cloning, blue-white selection, plasmid isolation. Transformation efficiency > 10 ⁸ . Ideal for cDNA generation and library construction. Genotype: F- mcrA Δ(mrr-hsdRMS-mcrBC) Φ80lacZΔM15 ΔlacX74 recA1 endA1 araD139 Δ(ara leu) 7697 galU galK rpsL nupG λ-	10 x 100 µl aliquots (10 tubes) 96 x 50 µl aliquots (12 x 8-tube strips)	T3019 T3020
XJa Autolysis [™] Mix & Q Competent Cells	JM109 with chromosomally inserted λ lysozyme gene inducible by arabinose. Genotype: <i>E. coli</i> K recA1 supE44 endA1 hsdR17 (r_k -, m_k ⁺) gyrA96 relA1 thi mcrAΔ(lac-proAB) Δ araB:: λ R, cat F'[traD36 proAB+ lacl ^q lacZ Δ M15]	10 x 100 µl aliquots (10 tubes)	T3021
XJb (DE3) Autolysis™ Mix & G Competent Cells	BL21(DE3) with chromosomally inserted λ lysozyme gene inducible by arabinose. DE3 lysogen encodes chromosomallyencoded T7 polymerase and is therefore a suitable host for expression of recombinant proteins under the control of the T7 promoter, such as in the pET system. Genotype: E. coli B F- ompT hsdSB(rB- mB-) gal dcm+ araB::R,cat λDE3	10 x 100 µl aliquots (10 tubes)	T3051

Other E. coli Related Products from Zymo Research

Rattler im Plating Beads flaming required. Spread cells evenly over the entire surface of the culture plate. 1 Bottle 5 Bottles 51001-5	Rattler™ Plating Beads	3	1 Bottle 5 Bottles	S1001 S1001-5
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^{*}Bulk quantities are available upon request. Please contact: <u>busdev@zymoresearch.com</u> or call 1-888-882-9682 for assistance.

What is Clean-Spin[™] Technology?

DNA PURIFICATION

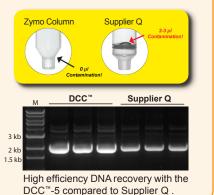
The spin columns from Zymo Research have been designed to ensure complete elution with no binding/wash buffer carryover. The result is ultra-pure inhibitor-free DNA and RNA.

Purify DNA from PCR & other sources

DNA Clean & Concentrator™ (DCC™)

- ✓ Recovery of ultra-pure DNA that is free of salts and contaminants.
- ✓ Small (≥6 µl) elution volume.
- ✓ DNA is ideal for ligation, PCR, Next-Gen sequencing, etc.

Product	Size (Cat. No.)
DNA Clean & Concentrator™-5	50 Preps. (D4013) 200 Preps. (D4014)
ZR-96 DNA Clean & Concentrator™-5	2 x 96 Preps. (D4023) 4 x 96 Preps. (D4024)
Genomic DNA Clean & Concentrator™	25 Preps. (D4010) 100 Preps. (D4011)



Boost DNA recoveries from agarose gels to >80%

Zymoclean™ Gel DNA Recovery

- ✓ Rapid (15 min.) recovery of ultra-pure DNA from agarose gels in ≥6 μl.
- ✓ Ultra-pure DNA ideal for DNA ligation, sequencing, etc.
- √ Format also available for large DNA >20 kb.

Product	Size (Cat. No.)
Zymoclean™ Gel DNA Recovery Kit	50 Preps. (D4001) 200 Preps. (D4002)
Zymoclean [™] Large Fragment DNA Recovery Kit	25 Preps. (D4045) 100 Preps. (D4046)

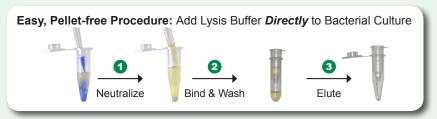


DNA fragments recovered from an agarose gel using the Zymoclean™ Gel DNA Recovery Kit. Lanes: M: DNA Ladder; 1-5: individual ladder DNA fragments.

Recover transfection-quality plasmid DNA directly from culture

Zyppy™ Plasmid Prep Kits

- ✓ The fastest, simplest method available for purifying high quality plasmid DNA from *E. coli*.
- ✓ Pellet-Free[™] procedure omits conventional cell-pelleting and resuspension steps.
- ✓ Transfection quality plasmid DNA directly from culture in under 15 minutes.



Product	Size (Cat. No.)
Zyppy™ Plasmid Miniprep Kit	50 Preps. (D4036) 100 Preps. (D4019) 400 Preps. (D4020) 800 Preps. (D4037)



The spin columns from Zymo Research have been designed to ensure complete elution with no binding/wash buffer carryover. The result is ultra-pure inhibitor-free DNA and RNA.

RNA PURIFICATION

Get RNA directly from TRIzol® without phase separation

Direct-zol™ RNA

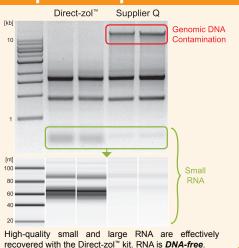
BIND

ELUTE

- ✓ For purification of high-quality small and large RNA directly from TRIzol®, TRI Reagent®, or similar.
- Bypasses phase separation and precipitation procedures allowing for unbiased recovery of miRNA

Product	Size (Cat. No.)
Direct-zol™ RNA MiniPrep	50 Preps. (R2050) 50 Preps. (R2051)* 200 Preps. (R2052) 200 Preps. (R2053)*
96-well and MagBead formats also available!	

DNase I included in all kits.

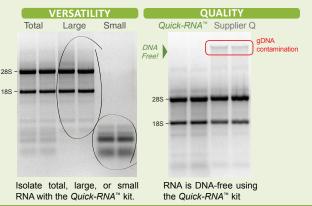


Isolate DNA-free RNA from 1 to 107 cells in minutes

Quick-RNA™

- Isolation of total, large, or small RNA You decide!
- Ultra clean, high-quality RNA from a single cell to 10⁷ cells.
- DNA-free RNA ideal for any downstream application DNase I included.

Product	Size (Cat. No.)
Quick-RNA™ MicroPrep	50 Preps. (R1050) 200 Preps. (R1051)
<i>Quick-RNA</i> ™ MiniPrep	50 Preps. (R1054) 200 Preps. (R1055)
ZR-96 Quick-RNA™	2 x 96 Preps. (R1052) 4 x 96 Preps. (R1053)



Purify RNA from enzymatic and labeling reactions in 5 minutes

RNA Clean & Concentrator™

- ✓ Recover ultra-pure RNA in small (≥6 μl) elution volumes.
- ✓ Compatible with TRIzol®, phenol, choloform, and RNase inhibitors (RNAlater®).
- ✓ RNA is ideal for RT-PCR, q-PCR, hybridization, arrays, RNA interference, etc.

Product	Size (Cat. No.)
RNA Clean & Concentrator™-5	50 Preps. (R1015) 200 Preps. (R1016)
RNA Clean & Concentrator™-25	50 Preps. (R1017) 100 Preps. (R1018)
ZR-96 RNA Clean & Concentrator™	2x96 well plates (R1080)
DNA-Free RNA Kit™	50 Preps. (R1013)



^{*} Supplied with TRI-Reagent®



The Beauty of Science is to Make Things Simple